THE REACTION OF DAMMARANE TRITERPENES WITH \underline{m} -Chloroperbenzoic acid¹)

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Abstract: Dammaran-20(\underline{S})-ol was treated with <u>m</u>-chloroperbenzoic acid (<u>m</u>CPBA) to give the corresponding 5¢-ol, 7 β -ol, 2¢-ol, 12 β -ol, 25-ol, and 5¢,25-diol. 20(\underline{S})-Hydroxydammaran-3 β -yl acetate was treated with <u>m</u>CPBA to afford the 5¢-ol, 7 β -ol, 12 β -ol, and 25-ol. Their structures were determined by spectroscopic methods.

Dammaranes are a common type of triterpene found in the plant kingdom, and are characteristic constituents of Ginseng²) and <u>Gynostemma pentaphyllum</u> Makino.³⁾ They are often hydroxylated at various positions besides C-3 and C-20. Tanaka and his co-workers reported that photolysis of $20(\underline{S})$ hydroxydammaran-3 α -yl p-nitrophenylacetate in Bu^tOH according to Breslow's procedure⁴) gave dammarane-3 α , 12 α , 20(\underline{S})-triol in 13 \underline{s} yield after hydrolysis.⁵) This was the first successful application of the remote oxidation to the triterpenes. No further reports of the application of this method to the triterpenes have appeared presumably due to the rather tedious steps required for derivation of the desired starting material.

We are interested in <u>in vitro</u> reactions which simulate the <u>in vivo</u> oxidation reactions observed in mammalian metabolism. We have been studying the structures of the metabolites obtained from rabbits upon feeding with various terpenoids.⁶⁾ The latter are generally excreted in hydroxylated form for detoxification.⁶⁾ We chose m-chloroperbenzoic acid (mCPBA) as the reagent for the hydroxylation of unactivated carbon atoms. Both concentrated peracids and radicals produced by photolysis of peroxides have previously found use but these reactions are, in principle, dangerous.⁷⁾ Deno <u>et al</u>. demonstrated that trifluoroperacetic acid can be used to hydroxylate aliphatic compounds, e.g. 2octanol to 2,7-octanediol. It is also useful for the cleavage of steroidal side chains.⁸⁾ mCPBA was first employed by Müller and Schneider for the introduction of a hydroxyl group at a bridgehead position of bicyclic compounds,⁹⁾ whilst Takaishi et al. later subjected polycyclic compounds¹⁰⁾ to the same procedure. We have recently applied this reaction to natural products, dammarane,¹⁾ lupane,¹¹⁾ and friedelane triterpenoids,¹¹⁾ cedrol,¹²⁾ and acyclic compounds¹³) and have found that hydroxylation occurs at not only tertiary but also secondary positions. We describe the details of the reactions of dammarane triterpenes with mCPBA and the assignments of the $^{13}\mathrm{C}$

NMR spectra of these products by use of INEPT techniques. Revision of the former assignments^{14,15}) is also discussed.

Dammaran-20(\underline{S})-ol (<u>1</u>) was treated with <u>mCPBA</u> (1.2 equiv.) in refluxing chloroform for 6 h. The reaction mixture was worked up and purified by column chromatography over silica gel to give six products (compounds I-VI) as well as the starting material (<u>1</u>) recovered in 77 % yield.

The ¹H NMR spectrum of compound I (<u>2</u>; Y. 59%¹⁶), $C_{30}H_{54}O_2$ [from high resolution mass, m/z 428.4044 (M-H₂O), and ¹³C NMR spectra], showed no peak between 3 and 5 ppm and in the ¹³C NMR spectrum thirty signals were observed, two of which were due to oxygen-bearing carbons [$\&_C$ 77.3 (s, C-5) and 75.4 (s,

C-20)]. The mass spectrum showed a molecular ion peak at m/z 446 and a characteristic peak at m/z 362 (Fig. 1).¹⁷⁾ These facts suggested that one hydroxyl group had been introduced at the C-5 position. ¹³C NMR signals having the similar chemical shifts to those of <u>1</u> are due to rings C and D and the side chain (Table 1). If the C-5 position is



substituted by a hydroxyl group, the chemical shifts of the carbons of rings A and B are well explained by suitable substituent effects. The stereochemistry of the C-5 position was determined by comparing the spectral data of $\underline{2}$ with those of <u>13</u> (<u>vide infra</u>). Compound I ($\underline{2}$) was thus determined to be dammarane- $5\alpha, 20(\underline{S})$ -diol.

In the ¹H NMR spectrum of compound II (3; Y. 2.18¹⁶), $C_{30}H_{54}O_2$ [HRMS m/z 428.4014 (M-H₂O) and ¹³C NMR] a double doublet centered at δ 3.80 (1H, J=11 and 5 Hz) was observed and the 13 C NMR spectrum indicated the presence of thirty carbons, two of which bore oxygen functions ($\delta_{\rm C}$ 75.3 (d, C-3) and 75.4 (s, C-20)]. From the coupling pattern of the methine proton, the hydroxyl group could be assigned to the 1 β , 7 β , or 15 α position. Of the five quaternary carbon atoms [$\delta_{\rm C}$ 75.4 (C-20), 49.7 (C-14), 46.2 (C-8), 37.3 (C-10), and 33.4 (C-4)) of 3, four had similar chemical shifts to those of dammaran- $20(\underline{S})$ -ol (<u>1</u>) [δ_{C} 75.3 (C-20), 50.4 (C-14), 40.6 (C-8), 37.5 (C-10), and 33.4 (C-4)], viz the carbon signal at $\delta_{\rm C}$ 40.6 assigned to C-8 has undergone a downfield shift of 5.6 ppm, indicating that the hydroxyl group was most probably in the 7β position. The secondary alcohol $(\underline{3})$ was oxidized to the corresponding ketone $(\underline{8})$, $C_{30}H_{52}O_2$ (m/z 444.3953), by Jones reagent. Methylene protons were observed at δ 2.50 (1H, t, J=15 Hz) and 2.20 (1H, dd, J=15 and 2.5 Hz) in the ¹H NMR spectrum of $\underline{8}$, suggesting the partial structure -CH₂-CO₋. From these results the secondary alcohol was determined to be dammarane- 7β ,20(S)-diol (3).

The third product, compound III ($\underline{4}$; Y. 1.3 $\16), $C_{30}H_{54}O_2$ [HRMS m/z 410.3920 (M-H₂O) and ¹³C NMR], was also a secondary alcohol from its ¹H NMR [$\frac{1}{6}$ 3.60 (1H, td, J=11 and 5 Hz)] and ¹³C NMR spectra [$\frac{1}{6}$ 74.4 (s, C-20) and 71.2 (d, C-12)]. The coupling pattern of the methine proton attached to the carbon bearing the newly introduced hydroxyl group indicated that the position of substitution was 6a, 11a, 12 β , or 16 β . Comparison of the ¹³C NMR chemical shifts (Table 1) with those of known dammarane triterpenes implied a 12 β -ol structure. Jones oxidation of $\underline{4}$ gave the ketone <u>9</u>, $C_{30}H_{52}O_2$ [m/z 426.3847 (M-



H₂O)], whose ¹H NMR spectrum showed a doublet at δ 2.85 (1H, J=10 Hz, H-13 β), a doublet of doublets at δ 2.30 (1H, J=14 and 5 Hz, H-11 α), and a triplet at δ 2.19 (1H, J=14 Hz, H-11 β), suggesting the partial structure -CH-CH₂-CO-CH-CH-. From these facts, compound III (<u>4</u>) was concluded to be dammarane-12,20(<u>5</u>)-diol (<u>4</u>).

The fourth product, compound IV ($\underline{5}$; Y. $28\16), $C_{30}H_{54}O_2$ [HRMS m/z 410.3927 (M-H₂O X 2) and ¹³C NMR], was easily assigned as dammarane-20(\underline{S}),25-diol ($\underline{5}$) from its mass spectrum [m/z 410 (M-H₂O X 2), 191, 145, 127 (base), and 59] and ¹H NMR spectrum (eight singlet methyls were observed at $|\delta$ 1.23, 1.23, 1.14, 0.96, 0.88, 0.85, 0.84, and 0.81).

A triplet of triplets observed at δ 3.89 (1H, J=11 and 4.5 Hz) in the ¹H NMR spectrum of the penultimate product, compound V ($\underline{6}$; Y. 4.2 $\16), C₃₀H₅₄O₂ [HRMS m/z 428.4209 (M-H₂O) and ¹³C NMR], indicated the presence of an equatorial secondary hydroxyl group with a methylene group on each side. The same pattern was observed in the ¹H NMR spectrum [δ 5.06 (1H, tt, J=10 and 6 Hz)] of its acetate (<u>10</u>). The position of substitution is limited only to 2a (equatorial). The corresponding ketone (<u>11</u>), C₃₀H₅₂O₂ [m/z 426.3853 (M-H₂O)], obtained by Jones oxidation showed four protons a to the carbonyl group [δ 2.39 (1H, dd, J=13 and 2 Hz), 2.28 (1H, d, J=13 Hz), 2.15 (1H, dd, J=13 and 2 Hz), and 1.98 (1H, d, J=13 Hz)]. These data were fully consistent with structure <u>6</u>, dammarane-2a, 20(<u>S</u>)-diol.

The most polar product, compound VI ($\underline{7}$; Y. 1.3 \times^{16}), $C_{30}H_{54}O_2$ [HRMS m/z 444.3999 (M-H₂O) and ¹³C NMR], showed eight singlet methyls in its ¹H NMR spectrum and fragment peaks at m/z 444 (M-H₂O), 378 (Scheme 4), 127, and 59 in its mass spectrum, suggesting the presence of a 25-hydroxyl group. In the ¹³C NMR spectrum thirty signals were detected, three of which corresponded to oxygenated carbons [δ_c 77.4 (s, C-5), 75.4 (s, C-20), and 71.1 (s, C-25)]. Comparison of the spectral data with those of <u>2</u> and <u>5</u> led directly to structure <u>7</u>.

We next carried out a similar reaction of $20(\underline{S})$ -hydroxydammaran-3 β -yl acetate (<u>12</u>) with <u>m</u>CPBA in chloroform. Work up and purification by column chromatography over silica gel and high performance liquid chromatography (HPLC) (μ -PORASIL, see Experimental) afforded four products, compound VII-X (<u>13-16</u>) as well as the recovered <u>12</u> (91% yield).

The mass fragmentation pattern (Scheme 4)¹⁷⁾ of the compound VII (<u>13</u>; Y. 16%¹⁶⁾), $C_{32}H_{56}O_4$ [HRMS m/z 486.4084 (M-H₂O) and ¹³C NMR], suggested a 5α-ol structure. As the methine proton attached to the carbon bearing the acetoxyl group appeared at δ 5.12 (dd, J=11 and 6 Hz), 0.64 ppm downfield of that of <u>12</u>, the hydroxyl group was determined to be 5α axial. The ¹³C NMR signals due to rings B, C, and D and the side chain of <u>13</u> were quite similar to those of <u>2</u> (<u>vide supra</u>). Thus the compound VII (<u>13</u>) was determined to be $5\alpha, 20(\underline{S})$ -dihydroxydammaran-3β-yl acetate.

The spectral data [m/z 486, 433, 419, 108, and 95; 6 4.48 (1H, dd, J=11 and 6 Hz), 3.79 (1H, dd, J=11 and 4 Hz), and 2.05 (3H, s); $\&_{\rm C}$ 171.0 (s, CO), 80.6 (d, C-3), 75.4 (s, C-20), and 70.9 (s, C-12)] of compound VIII (<u>14</u>; Y. 1.6%¹⁶), $C_{32}H_{56}O_4$ [HRMS m/z 486.4087 (M-H₂O) and ¹³C NMR], are very well explained in terms of the 3 β -acetate of compound II (<u>3</u>) and hence compound VIII (<u>14</u>) should be formulated as 7β ,20(<u>S</u>)-dihydroxydammaran-3 β -yl acetate.

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In the ¹H NMR spectrum of compound IX (<u>15;</u> Y. 1.6%¹⁶), $C_{32}H_{56}O_4$ [HRMS m/z 468.3956 (M-H₂O X 2) and ¹³C NMR], a triplet of doublets (J=10 and 5 Hz) was observed at δ ; 3.60, which was exactly the same position and pattern as that of <u>4</u>. The data were completely identical with those of authentic 12 β ,20(<u>\$</u>)dihydroxydammaran-3 β -yl acetate (<u>15</u>).¹⁴) As this is an acetate of a typical sapogenin of Ginseng, this constitutes a one step synthesis of such types from a 12-deoxy compound.

The last compound X (<u>16</u>; Y. 40.3 $\16),¹⁸) $C_{32}H_{56}O_4$ [HRMS m/z 486.4112 (M-H₂O) and ¹³C NMR], exhibited eight singlet methyls (δ 1.23, 1.23, 1.14, 0.96, 0.88, 0.87, 0.85, and 0.85) in its ¹H NMR spectrum and characteristic fragmentations [m/z 468 (M-H₂O X 2), 191, 189, 145, 127, 109, and 59] in its mass spectrum. Hence it was determined to be 20(<u>S</u>),25-dihydroxydammaran-38-yl acetate (<u>16</u>).

Thus the hydroxylation reaction reported here takes place by an attack on unactivated carbon atoms of the triterpene framework. The mechanism is presumably a radical process as recently reported by Fossy <u>et</u>. <u>al</u>.¹⁹⁾ The order of reactivity of unactivated carbon atoms is tertiary > secondary > primary. Hydroxylation of the 25 position is the major reaction because it is a tertiary position and is not subject to steric hindrance. The 5 α -ol is another major product of both reactions, even though the 5-position would appear to be subject to somewhat more steric hindrance. The other products are all equatorial alcohols (2 α , 7 β , and 12 β -ol), showing that equatorial alcohols are formed much more readily than the corresponding axial alcohols. Although the reaction site cannot fully be predicted at this stage, the oxidation reaction with <u>mCPBA</u> is quite useful for introduction of a functional group at an unactivated carbon atom.

 13 C NMR Tanaka and his co-workers assigned the 13 C NMR spectra of a number of dammarane triterpenes and these results were later used to assign the 13 C NMR spectrum of $1.^{14,15}$) However, measurement of 13 C NMR spectrum of 1 using the INEPT pulse sequence has revealed that the previously determined multiplicities of two resonances are incorrect. Thus the signal at $\delta_{\rm C}$ 24.8 is due to a methylene carbon whilst that at $\delta_{\rm C}$ 25.6 results from a methyl group (Fig. 2). It is probable that the assignments of Tanaka are also in need of revision. In order to ensure the correctness of other assignments, the 13 C NMR spectra of several derivatives in hand were determined and the multiplicities confirmed by use of INEPT pulse sequence. In the case of triterpenes most 13 C NMR signals appear in the highly congested region between 10 and 50 ppm and it is hence difficult to determine the multiplicities from the SFORD spectrum. The INEPT or DEPT pulse sequences are the best choice for such compounds.

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	<u> </u>	<u> </u>	3	4	2	<u>₽</u>	<u></u>	<u>13</u>	14	15	<u></u>	
1	40.6	32.9	40.5	40.5	40.7	49.9	32.8	32.1	36.9	37.9	38.8	
2	18.7	17.9	18.7	18.7*	18.7	65.3	17.9	23.6	21.4	23.7	23.7	
3	42.2	36.2	41.9	42.1	42.2	51.2	36.1	77.2	80.6	80.8	81.0	
4	33.4	38.5	33.2	33.3	33.4	39.7	38.4	39.3	38.5	38.6	38.0	
5	57.0	77.3	54.3	57.0	57.0	56.4	77.4	78.9	53.3	55.9	56.0	
6	18.7	23.0	28.2	18.6*	18.7	18.3	23.0	23.4	27.7	18.2	18.2	
7	35.3	28.7	75.3	34.8	35.3	35.0	28.6	28.4	75.1	34.8	35.2	
8	40.6	40.9	46.2	39.9	40.7	41.0	41.0	40.7	46.0	39.8	40.4	
9	50.8	42.0	48.8	50.2	50.8	50.7	41.9	42.0	48.7	50.0	50.6	
10	37.5	39.6	37.3	37.4	37.5	42.1	39.6	43.1	37.7	37.0	37.1	
11	21.5	21.5	21.2	31.2	21.5	21.5	21.5	21.7	21.3	31.2	21.6	
12	24.8	24.8	25.3	71.2	24.9	24.8	24.8	24.8	23.7	70.9	24.9	
13	42.2	42.3	43.0	47.7	42.4	42.1	42.3	42.4	43.0	47.8	42.3	
14	50.4	50.9	49.7	51.7	50.4	50.4	50.8	50.8	49.7	51.6	50.3	
15	31.2	31.3	34.7	30.9	31.2	31.1	31.3	31.3	34.7	31.0	31.2	
16	27.6	27.8	27.4	26.6	27.7	27.5	27.8	27.8	27.3	26.5	27.5	
17	49.8	49.8	50.4	53.5	50.0	49.7	49.9	49.8	50.1	53.5	49.9	
18	16.2	16.7	9.8	16.1	16.2	15.6	16.7	16.6	9.7	16.2	16.3	
19	15.6	19.2	16.1	15.6	15.6	17.3	19.2	19.3	16.2*	15.7	15.5	
20	75.3	75.4	75.4	74.4	75.4	75.4	75.5	75.4	75.4	74.5	75.4	
21	25.6	25.6	25.7	27.1	25.6	25.6	25.5	25.7	25.8	27.2	25.6	
22	41.1	41.1	40.9	35.1	41.1	40.5	40.9	41.2	40.9	35.0	41.1	
23	21.5	21.5	21.5	21.3	18.5	21.6	18.5	21.5	21.5	21.3	18.5	
24	39.8	39.8	39.7	39.8	44.6	39.1	44.5	39.8	39.7	39.8	44.5	
25	28.0	28.1	28.0	28.2	71.1	28.0	71.1	28.1	28.0	28.2	71.0	
26	22.7	22.6	22.6*	22.6+	29.3*	22.6*	29.3*	22.7*	22.6+	22.6*	29.3*	
27	22./	22.6	22.7*	22.7+	29.5*	22.7*	29.4*	22.8*	22.7+	22.7*	29.5*	
28	33.4	27.9	33.3	33.4	33.4	33.5	27.9	22.6	28.0	28.0	28.0	
29	21.5	23.9	21.5	21.5	21.6	42.4	23.9	18.5	16.5	16.5	16.5	
30	16.5	15./	10.4	16.9	10.0	10.4	15./	15./	10.3*	10.8	10.5	
00								170.9	171.0	1/1.0	1/1.0	
CHo								21.3	25.3	21.3	21.3	

Table 1. ¹³C NMR data of dammarane derivatives

*,+ Assignments may be reversed.



Fig. 2. (a) Complete ¹H decoupled spectrum (100 MHz) of dammaran-20(\underline{S})-ol (<u>1</u>). About 20 mg/0.5 ml CDCl₃ solution, 100 times accumulation, pulse width 4 μ s (ca. 40°), pulse delay 2 sec, data points 16 K, frequency 20000 Hz. (b) INEPT spectrum with PI1(=1/4J)=2 ms, PI2(=3/4J)=6 ms. (c) INEPT spectrum with PI1(=1/4J)=2 ms.

EXPERIMENTAL

All melting points were measured on a Yanagimoto microscope hot plate and uncorrected. IR spectra were measured with a Shimadzu IR-27G trophotometer. ¹H and ¹³C NMR spectra were obtained with a JBOL JMN-GX400 are spectrophotometer. spectrophotometer. In and C NEW spectra were obtained with a check on a spectrometer in CDCl₃ unless otherwise stated. Mass spectra were taken on a Shimadzu LKB-9000B or JEOL JMS D-300. Thin layer chromatography (TLC) was performed on Silica gel 60 F_{254} (Merck). Silica gel 60 (70-230 mesh, Merck) was used for column chromatography. <u>mCPBA was purchased from Nakarai Chemical Co.</u> Ltd. and used without further purification.

used for column chromatography: mCPBA was purchased from Nakarai Chemical Co. Ltd. and used without further purification. Oxidation Reaction of Dammaran-20(§)-ol (1) with mCPBA To a solution of dammaran-20(§)-ol (1) (1.0 g) in CHCl₁ (30 ml) was added mCPBA (450 mg, 1.2 g). The mixture was attrired at 65-75'C for 6h and washed successively with 58 Na 250₃ ag, 5% NaHCO₃ ag, and brine. The dried solution was evaporated in vacuo to give the residue (1.23 g). The product mixture was chromatographed on silica gel, using hexane-Et₂0 and benzene-EtOAc solvent systems for elution. The products were further pulified by prep. HPLC (u-PORASIL, 15 % EtOAc-hexane) to afford danmaran-20(§)-ol (1) (140 mg), dammarane-320(§)-diol (2) (140 mg), dammarane-20(2)-diol (3) (5 mg), dammarane-220(5)-diol (2) (10 mg), and dammarane-53,20(§)-diol (5) (66 mg), dammarane-220,3)-diol (5) (10 mg), and dammarane-53,20(5)-diol (5) (15 mg), dammarane-220,3)-diol (5) (10 mg), and dammarane-53,20(5)-diol (2) (19 mg) 16°C; IR λ_{max} : 3620, 3460 (OH) cm⁻¹; ¹H NMR: 6 0.87 (3H, s), 0.98 (3H, d), J=7 Hz), 0.99 (3H, d, J=7 Hz), 0.96 (3H, s), 0.96 (3H, s), 1.00 (3H, s), 1.10 (3H, s), 1.10 (3H, s), 2.1-CH₃); MS MMR (Cp₆): 6 0.86 (3H, s), 0.92 (3H, d), J=7 Hz), 0.93 (3H, d, J=7 Hz), 0.94 (3H, s), 0.96 (3H, s), 0.93 (3H, d), J=7 Hz), 0.94 (3H, s), 0.95 (3H, s), 0.97 (3H, s), 1.13 (3H, s), 2.1-CH₃); MS MKR (Cp₆): 6 0.82 (3H, s), 0.96 (3H, s), 0.87 (3H, s), 0.87 (3H, s), 0.87 (3H, s), 0.95 (3H, s), 0.97 (3H, s), 0.81 (3H, 30, 285, 247, 123, 109, 95, 69 (base); HRMS: Found m/z 428.4014 (M-H₂O)², Calcd for C₃₀H₂₅₀O 428.4018. Damarane-128,20(S)-diol (4): 0:11; IR λ_{max} : 3600, 3450 (0H) cm⁻¹; ¹ H NMR: 16 0.81 (3H, s), 0.87 (3H, s), 0.88 (6H, d, J=1, 5 Hz, 7-Ha); MS m/z: 428 (M-H₂O)², 410, 361, 343, 318, 300, 285, 247, 123, 109, 95, 69 (base); HRMS: Found m/z 428.4014 (M-H₂O)², Calcd for C₃₀H₅₀O 428.4019. Damarane-128,20(S)-diol (4): 0:11; IR λ_{max} : 3600, 3450 (0H) cm⁻¹; ¹ H NMR: 16 0.81 (0.84, 0.85, 0

Oxidation of Diol 3 The diol 3 (2.2 mg) in acetone (0.6 ml) was oxidized with Jones reagent (2 drops) at rt for 30 min to give 7-oxpdammaran-20(S)-ol (8) (1.9 mg): Oil; IR λ_{max} : 3400 (OH), 1700 (C=O) cm⁻¹; ¹H NMR: δ 0.83 (3H, s), 0.84 (3H, s), 0.88 (6H, d, J=6.5 Hz, 26-CH₃, 27-CH₃), 0.89 (3H, s), 1.00 (3H, s), 1.13 (3H, s), 1.16 (3H, s), 2.20 (1H, dd, J=15, 2.5 Hz, 6-Ha), 2.50 (1H, t, J=15 Hz, 6-Hb); ¹H NMR (C₆D₆): δ 0.67 (3H, s), 0.71 (3H, s), 0.77 (3H, s), 0.92 (3H, d, J=6.5 Hz), 0.93 (3H, d, J=6.5 Hz), 1.04 (3H, s), 1.06 (3H, s), 1.08 (3H, s); MS m/z: 426 (M-H₂O)⁺, 410, 359, 316, 248, 220, 123, 95, 69 (base); HRMS: Found m/z 444.3953 (M⁺), Calcd for C₃₀H₅₂O₂ 444.3965.

Oxidation of Diol 4 To a stirred solution of the diol 4 (1.0 mg) in acetone (0.4 ml) was added Jones reagent (2 drops) at rt. The mixture was stirred for 30 min and acetone was removed. Work up as usual afforded 12-oxodammaran- $20(\underline{S})-o1$ ($\underline{9}$) (0.7 mg): Oil: IR $|\lambda_{max}$: 3400 (OH), 1700 (C=O) cm⁻¹; ¹H NMR: $|\delta$ 0.81 (3H, s), 0.84 (3H, s), 0.87 (3H, s), 0.87 (3H, d, J=7 Hz), 0.88 (3H, d, J=7 Hz), 0.93 (3H, s), 1.10 (3H, s), 1.17 (3H, s, 21-CH₃), 2.19 (1H, t, J=14 Hz, 11-Hb), 2.30 (1H, dd, J=14, 5 "z, 11-Ha), 2.40 (1H, td, J=10, 7 Hz, 17-H), 2.85 (1H, d, J=10 Hz, 13-H); MS m/z: 426 $(M-H_2O)^+$, 411, 359, 316, 234, 191, 124 (base); HRMS: Found m/z 426.3847 $(M-H_2O)^+$, Calcd for $C_{30}H_{50}O$ 426.3859.

Acetylation of Diol <u>6</u> The diol <u>6</u> (8.0 mg) was treated with acetic anhydride (1 ml) in pyridine (1 ml) and worked up as usual to give $20(\underline{S})$ -hydroxydammaran-2 α -yl acetate (<u>10</u>) (8 mg): mp 144°C (EtOAc); IR λ_{max}^{i} : 3450 (OH), 1720 (C=O) cm⁻¹; H NMR: δ_{1}^{2} .02 (3H, s, OCOCH₃), 5.06 (1H, tt, J=10, 6 Hz, 2-H β); MS m/z: 470 (M-H₂O)⁺, 410, 403, 300, 285, 191, 189, 129 (base); HRMS: Found m/z 470.4110 (M-H₂O)⁺, Calcd for C₃₂H₅₄O₂ 470.4122.

Oxidation of Diol 6 The diol 6 (9.0 mg) in acetone (1.6 ml) was oxidized with Jones reagent (4 drops) at rt for 1h to give 2-oxodammaran-20(\underline{S})-ol (11) (8.4 mg): Oil; IR $\lambda|_{max}$: 3450 (OH), 1700 (C=O) cm⁻¹; ¹H NMR: 6 0.86 (3H, \underline{s}), 0.88 (6H, d, J=7 Hz), 0.89 (3H, s), 0.92 (3H, s), 0.97 (3H, s), 1.06 (3H, s), 1.13 (3H, s), 1.98 (1H, d, J=13 Hz), 2.15 (1H, dd, J=13, 2 Hz); ¹H NMR (C₆D₆): 6 0.78 (6H, s), 0.79 (3H, s), 0.84 (3H, s), 0.86 (3H, s), 0.93 (3H, d, J=7 Hz), 0.94 (3H, d, J=7 Hz), 1.09 (3H, s), 1.57 (1H, d, J=13 Hz), 1.96 (1H, d, J=13 Hz), 2.13 (1H, dd, J=13, 2 Hz), 2.37 (1H, dd, J=13, 2 Hz); MS m/z: 426 (M-H₂O)⁺, 316, 217, 205, 95, 69 (base); HRMS: Found m/z 426.3853 (M-H₂O)⁺, Calcd for C₂₀H₅O 426.3860. for C₃₀H₅₀O 426.3860.

Oxidation Reaction of $20(\underline{S})$ -Hydroxydammaran-3 β -yl acetate (<u>12</u>) with mCPBA <u>mCPBA</u> (843 mg, 1.2 eq) was added to a solution of $20(\underline{S})$ -hydroxydammaran-3 β -yl acetate (<u>12</u>) (1.44 g) in CHCl₃ (45 ml) in one portion and the mixture was refluxed for 6h. The reaction mixture was worked up as usual. The residue (1.65 g) was purified by silica gel column chromatography (benzene-EtOAc) to give <u>12</u> (1.32 g), a mixture of <u>13</u>, <u>14</u>, <u>15</u>, and <u>16</u> (50 mg). The mixture of <u>13</u>, <u>14</u>, and <u>15</u> was further purified by prep. HPLC (μ +PORASIL, 5% EtOAc-hexane and 20% EtOAc-hexane) to give 5α , $20(\underline{S})$ -dihydroxydammaran-3 β -yl acetate (13) (20

14, and 15 was further purified by prep. HPLC (μ -PORASIL, 5% EtOAc-hexane and 20% EtOAc-hexane) to give 5%,20(S)-dihydroxydammaran-3&yl acetate (13) (20 mg), 7 β ,20(S)-dihydroxydammaran-3&yl acetate (14) (2 mg), and 12 β ,20(S)-dihydroxydammaran-3&yl acetate (15) (2 mg). 5%,20(S)-Dammaran-3&yl acetate (15) (2 mg). 5%,20(S)-Dammaran-3&yl acetate (13): mp 194°C; IR λ_{max} : 3600, 3450 (OH), 1725, 1255 (OCO) cm⁻¹; ¹H NMR: δ 0.88 (6H, d, J=7 Hz, Y26-CH₃, 27-CH₃), 0.91 (3H, s), 0.94 (3H, s), 0.96 (3H, s), 1.00 (3H, s), 1.02 (3H, s), 1.13 (3H, s, 21-CH₃), 2.03 (3H, s, OCOCH₂), 5.12 (1H, dd, J=11, 6 Hz, H-3 α); MS m/z: 486 (M-H₂O)⁺, 468, 426, 393, 362 (base), 344, 259, 95, 69; HRMS: Found m/z 486.4084 (M-H₂O)⁺, Calcd for C₃₂H₅Q₃ 486.4072. 7 β ,20(S)-Dammaran-3B-yI acetate (14): 0il; IR λ_{max} : 3450 (OH), 1720, 1250 (OCO) cm⁻¹; ¹H NMR: δ 0.86 (3H, s), 0.87 (6H, s), 0.88 (6H, d, J=7 Hz), 0.97 (3H, s), 1.14 (3H, s, 21-CH₃), 2.05 (3H, co COCH₃), 3.79 (1H, dd, J=11, 6 Hz, H-3 α); MS m/z: 486 (M-H₂O)⁺, Calcd for C₃₂H₅Q₃ 486.4072. 7(J), 4.48 (1H, dd, J=11, 6 Hz, H-3 α); MS m/z: 486 (M-H₂O)⁺, 433, 419, 343, 303, 298, 283, 249, 189, 129, 108, 95 (base), 69, 43; HRMS: Found m/z 486.4087 (M-H₂O)⁺, Calcd for C₃₂H₅Q₃ 486.4072. 12 β ,20(S)-Dihydroxydammaran-3 β -yI acetate (15): 0il: IR λ_{max} : 3350 (OH), 1725, 1250 (OCO) cm⁻¹; ¹H NMR: δ 0.86 (3H, s), 0.87 (3H, d, J=7 Hz), 0.88 (3H, d, J=7 Hz), 0.89 (3H, s), 0.91 (3H, s), 0.99 (3H, s), 1.19 (3H, s, 21-CH₃), 2.05 (3H, s), 0.020 (cm⁻¹; IH NMR: δ 0.86 (3H, s), 0.87 (3H, d, J=7 Hz), 0.88 (3H, d, J=10, 5 Hz, H-120); 4.48 (1B, dd, J=11, 6 Hz, H-3 α); MS m/z: 468 (M-H₂O x 2)⁺ Calcd for C₃₂H₅O₂ 468.3956 (M-H₂O x 2)⁺ Calcd for C₃₂H₅O₂ 468.3957. 20(S), 25-Dihydroxydam

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REFERENCES AND NOTES

- A part of this paper was published in a preliminary form: M. Tori, R. Matsuda, Y. Asakawa, Tetrahedron Lett., <u>26</u>, 227 (1985).
 S. Shibata, M. Fujita, S. Itokawa, O. Tanaka, T. Ishii, Chem. Pharm. Bull.,
- <u>11</u>, 759 (1963). 3) T. Takemoto, S. Arihara, T. Nakajima, M. Okuhira, Yakugaku Zasshi, <u>103</u>, 173
- (1983).
- 4) R. Breslow, Chem. Soc. Rev., <u>1</u>, 553 (1972). 5) R. Kasai, K. Shinzo, O. Tanaka, K. Kawai, Chem. Pharm. Bull., <u>25</u>, 1213 (1974).
- 6) T. Ishida, Y. Asakawa, T. Takemoto, J. Pharm. Sci., <u>71</u>, 965 (1982) and references cited therein.
 7) <u>e. q</u>. D. L. Heywood, B. Phillips, H. A. Stansbury, Jr., J. Org. Chem., <u>61</u>, 281 (1961).
- 281 (1961).
 8) N. C. Deno, L. A. Messer, Chem. Commun., <u>1976</u>, 1051; N. C. Deno, M. D. Meyer, J. Org. Chem., <u>44</u>, 3383 (1979).
 9) W. Müller, H. -J. Schneider, Angew. Chem., Int. Ed. Engl., <u>18</u>, 407 (1979).
 10) N. Takaishi, Y. Fujikura, Y. Inamoto, Synthesis, <u>1983</u>, 293.
 11) M. Tori, R. Matsuda, Y. Asakawa, Chem. Lett., <u>1985</u>, 167.
 12) M. Tori, R. Matsuda, Y. Asakawa, Bull. Chem. Soc. Jpn., <u>58</u>, 2523 (1985).
 13) M. Tori, M. Sono, Y. Asakawa, Bull. Chem. Soc. Jpn., <u>58</u>, 2669 (1985).
 14) J. Asakawa, R. Kasai, K. Yamasaki, O. Tanaka, Tetrahedron, <u>33</u>, 1935 (1977).
 15) M. Tori, T. Tsuyuki, T. Takahashi, Bull. Chem. Soc. Jpn., <u>50</u>, 3349 (1977).
 16) Yields shown are isolation yields based on consumed starting material.
 17) T. R. Govindachari, N. Viswanathan, P. A. Mohamed, Tetrahedron, <u>27</u>, 4991 (1971).

- (1971).
- 18) T. Ikeda, K. Kitao, Mokuzai Gakkaishi, <u>20</u>, 460 (1974).
 19) J. Fossy, D. Lefort, M. Massoudi, J. -Y. Nedelec, J. Sorba, Can. J. Chem., <u>63</u>, 678 (1985).